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Chiral syntheses of 6'-β-fluoroaristeromycin, 6'-β-fluoro-5'-noraristeromycin and aristeromycin

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Abstract—Carbocyclic nucleosides substituted at the C-6' position are receiving increasing attention. Chiral synthetic accessibility to the biologically promising 6'- β -fluoroaristeromycin is lacking. Its preparation and that of the 5'-nor analogue are described. Along the way, a new method to aristeromycin arose as an outgrowth of a requisite structure proof. © 2005 Elsevier Ltd. All rights reserved.

In using the nucleoside scaffold for medicinal agent and biochemical mechanistic investigations, the C-6' site of the carbocyclic nucleoside framework¹ (see aristeromycin, 1, Fig. 1) offers a center for modification² not available in the more common ribofuranosyl based nucleosides. Because of the promising properties synthetically derived fluorinated carbocyclic nucleosides have displayed,³ we were attracted to (\pm) -6'- β -fluoro-aristeromycin (2) as a representative of the C-6' substituted carbocyclic series that has shown promising potential as an antiviral (orthopox) agent.^{2b} This latter property, however, has not been fully developed. In that direction, a chiral synthesis of the enantiomer of 2 (i.e., 3) resembling the parent aristeromycin (1) has been completed in our laboratory. This undertaking concurrently prompted an interest in seeking access to the 5'noraristeromycin fluoro analogue 4 (and its enantiomer, 5) as an extension of our work with 5'-noraristeromycin (6)⁴ as a source of new antiviral candidates. Accompanying these investigations was a convenient preparation of aristeromycin (1). The results of these efforts are communicated here.

In designing synthetic procedures to carbocyclic nucleosides, our laboratory has always sought a common starting point. In that regard, the approach to 3-5 (that also opened a means to 1) was built upon (+)-(1R,4S)-4-

hydroxy-2-cyclopenten-1-yl acetate (7),⁵ which has served us well in our studies.⁶ From this, two guiding features followed in the plan to 3: (i) an enantiospecific synthesis of 8 (Scheme 1) for conversion into 3 following the literature precedence^{2b} and (ii) application of our recent report employing a stereo-controlled epoxide formation with subsequent nucleophilic opening as a means to polyfunctionalized cyclopentanes.⁷

To begin, 10, obtained from 7,8 was subjected to a Tamao oxidation8 to 11 whose primary alcohol center was protected as its *tert*-butyldiphenylsilyl derivative 12 (Scheme 1). Mitsunobu inversion of the secondary alcohol of 12 followed by glycolization, *iso* propylidene protection and ammonolysis (to remove the acetate from the inverted alcohol) formed 13. Confirmation of the functional group stereochemistry depicted in 13 was accomplished by transforming it into 1 by, first, pyridinium chlorochromate oxidation followed by Luche reduction to 14. Mitsunobu coupling of 14 with 6-chloropurine and subsequent desilylation led to 15. Ammonolysis of 15 and, then, acidic deprotection yielded 1 whose spectral properties were identical to those reported by us9 and others in the literature. 10

Confident of the structure of 13, and after evaluating a number of procedures, a Mitsunobu-type elimination reaction (Scheme 1, step h) resulted in the conversion of 13 into 16. Drawing upon our previous results, conversion of 16 into 8 required primary alcohol protection with a benzyl group. Thus, desilylation/benzylation led to 9. Subjecting 9 to the sequence of (i) deisopropylidenation, (ii) epoxidation, (iii) sodium azide ring opening,

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Figure 1.

Scheme 1. Reagents and conditions: (a) Ref. 8; (b), KF, H₂O₂, MeOH/THF/H₂O, 87%; (c) TBDPSCl, imidazole, CH₂Cl₂, 70%; (d) (i) TPP, DIAD, AcOH, THF; (ii) NMO, OsO₄, acetone/THF/H₂O; (iii) Me₂C(OMe)₂, acetone, pTSA, 61% for three steps; (iv) NH₃, MeOH, 86%; (e) (i) PCC, Celite, CH₂Cl₂; (ii) NaBH₄, CeCl₃·7H₂O, MeOH, 90% for two steps; (f) (i) TPP, DIAD, 6-chloropurine, THF; (ii) TBAF, THF, 71% for two steps; (g) Ref. 9b; (h) TPP, DIAD, toluene, 80 °C, 10 h, 82%; (i) (i) TBAF, THF, 95%; (ii) BnBr, NaH, DMF, 91%; (j) (i) HCl, MeOH; (ii) mCPBA, CH₂Cl₂; (iii) NaN₃, DMF/H₂O, (iv) Me₂C(OMe)₂, acetone, pTSA, 63% for 4 steps; k, (i) Tf₂O, pyridine, CH₂Cl₂; (ii) TASF, THF, 85% for two steps^{2b}; (l) (i) Lindlar catalyst, H₂, MeOH; (ii) 5-amino-4,6-dichloropyrimidine, Et₃N, 1-BuOH, 105 °C; (iii) (EtO)₂CHOAc, reflux, 38% for three steps;^{2b} (m) (i) NH₃, MeOH, 69%; (ii) Pd(OH)₂/C, cyclohexene, EtOH, reflux; (iii) 1 N HCl, MeOH, 70 °C, 86% for two steps.^{2b}

and (iv) *iso*propylidenation gave **8**¹⁰ in a 4:1 ratio along with its isomer **i**. The structure of **8** was confirmed by comparing its NMR spectral data with the reported racemic **8** prepared in another way.¹⁰

Alcohol 8 was then converted into 3^{11} by utilizing a reported procedure^{2b} as delineated in Scheme 1.

With the intent of utilizing a similar epoxide-sodium azide ring opening path to 4 and 5, silyl protection of 7–19^{6b} was followed by glycolization and, then, *iso* propylidenation and ammonolysis of the resultant acetate to afford 20.¹² Subjecting 20 to the Mitsunobu process

used with 13 followed by desilylation gave 21, which was converted into the benzyl derivative 22. Following the same series of steps used for the 9 to 8 process, 22 yielded 23 (Scheme 2). To confirm the stereochemical orientation of 23, it was converted into the known 24¹³: (i) iodination with inversion to 25, (ii) removal of the iodine concurrently with reduction of the azide, (iii) reaction of the free amino with phthalic anhydride, and (iv) subsequent debenzylation with deisopropylidenation.

Returning to the preparation of 4, fluorinative inversion of 23 to 26 was followed by azide reduction to the primary amine, which lent itself to the standard stepwise

RO OAC
$$\frac{b}{\text{on 19}}$$
 TBSO OH $\frac{b}{\text{on 19}}$ OH $\frac{b}{\text{on 22}}$ OH $\frac{b}{\text{on 22$

Scheme 2. Reagents and conditions: (a) TBSCl, imidazole, CH₂Cl₂, 91%; (b) (i) OsO₄, NMO, acetone/H₂O; (ii) Me₂C(OMe)₂, acetone, *p*TSA, 90% for two steps; (iii) NH₃, MeOH, 85%; (c) (i) TPP, DIAD, toluene, 80 °C, 93%; (ii) TBAF, THF, 95%; (d) BnBr, NaH, DMF, 97%; (e) (i) HCl, MeOH; (ii) *m*CPBA, CH₂Cl₂; (iii) NaN₃, DMF/H₂O; (iv) Me₂C(OMe)₂, acetone, *p*TSA, 59% for four steps; (f) (i) Tf₂O, pyridine, CH₂Cl₂; (ii) LiI, THF 92% for two steps; (g) (i) H₂, Pd/C, MeOH, 90%; (ii) phthalic anhydride, Ac₂O, pyridine; (iii) Pd(OH)₂/C, cyclohexene, EtOH, 56% for two steps; (h) (i) Tf₂O, pyridine, CH₂Cl₂; (ii) TASF, THF, 77% for two steps; (i) (i) Lindlar catalyst, H₂, MeOH; (ii) 5-amino-4,6-dichloropyrimidine, Et₃N, 1-BuOH; (iii) (EtO)₂CHOAc, reflux, 35% for three steps; (j) (i) NH₃, MeOH, 75%; (ii) Pd(OH)₂/C, cyclohexene, EtOH, 100%; (iii) 1 N HCl, MeOH, 96%; (k) Ref. 17; (l) Refs. 5c and 7; (m) steps (d), (e), (h), (i), and (j).

purine construction process^{2b,14} (Scheme 2, step i) to provide **27**. Ammonolysis, debenzylation and then de*iso* propylidenation yielded **4**.¹⁵

Enantiomer 5^{16} was prepared from 28 using a procedure similar to that for obtaining 4.

The biological analysis of 3–5 is underway and will be presented in the full paper on this class of nucleoside derivatives.

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References and notes

- (a) Schneller, S. W. Curr. Top. Med. Chem. 2002, 2, 1087–1092;
 (b) Rodriguez, J. B.; Comin, M. J. Mini Rev. Med. Chem. 2003, 3, 95–114.
- (a) Cheikh, A. B.; Craine, L. E.; Recher, S. G.; Zemlicka, J. J. Org. Chem. 1988, 53, 929–936; (b) Madhavan, G. V. B.; McGee, D. P. C.; Rydzewski, R. M.; Boehme, R.; Martin, J. C.; Prisbe, E. J. J. Med. Chem. 1988, 31, 1798–1804; (c) Hong, J. H.; Oh, C. H.; Cho, J. H. Tetrahedron 2003, 59, 6103–6108; (d) Kim, J. W.; Choi, B. G.; Hong, J. H. Bull. Korean Chem. Soc. 2004, 25, 1812–1816; (e) Ruediger, E.; Martel, A.; Meanwell, N.; Solomon, C.;

- Turmel, B. *Tetrahedron Lett.* **2004**, *45*, 739–742; (f) Takagi, C.; Sukeda, M.; Kim, H. S.; Wataya, Y.; Yabe, S.; Kitade, Y.; Matsuda, A.; Shuto, S. *Org. Biomol. Chem.* **2005**, *3*, 1245–1251.
- 3. (a) Borthwick, A. D.; Kirk, B. E.; Biggadike, K.; Exall, A. M.; Butt, S.; Roberts, S. M.; Knight, D. J.; Coats, J. A. V.; Ryan, D. M. J. Med. Chem. 1991, 34, 907-914; (b) Nakayama, T.; Matsumura, Y.; Morizawa, Y.; Yasuda, A.; Uchida, K.; Takase, H.; Murakami, Y.; Atarashi, S.; Ikeuchi, T.; Osada, Y. Chem. Pharm. Bull. 1994, 42, 183-187; (c) Jeong, L. S.; Moon, H. R.; Park, J. G.; Shin, D. H.; Choi, W. J.; Lee, K. M.; Kim, H. O.; Chun, M. W.; Kim, H. D.; Kim, J. H. Nucleosides Nucleotides Nucleic Acids 2003, 22, 589-592; (d) Yang, Y. Y.; Meng, W. D.; Qing, F. L. Org. Lett. 2004, 6, 4257-4259; (e) Rosen, T. C.; De Clercq, E.; Balzarini, J.; Haufe, G. Org. Biomol. Chem. 2004, 2, 229–237; (f) Kim, H. O.; Yoo, S. J.; Ahn, H. S.; Choi, W. J.; Moon, H. R.; Lee, K. M.; Chun, M. W.; Jeong, L. S. Bioorg. Med. Chem. Lett. 2004, 14, 2091-2093; (g) Moon, H. R.; Lee, H. J.; Kim, H. R.; Lee, K. M.; Lee, S. K.; Kim, H. O.; Chun, M. W.; Jeong, L. S. Bioorg. Med. Chem. Lett. 2004, 14, 5641-5644.
- 4. For a leading reference see: Yin, X. Q.; Schneller, S. W. *Tetrahedron* **2005**, *61*, 1839–1843.
- (a) Laumen, K.; Reimerdes, E. H.; Schneider, M.; Gorisch, H. Tetrahedron Lett. 1985, 26, 407–410; (b) Laumen, K.; Schneider, M. Tetrahedron Lett. 1984, 25, 5875–5878; (c) Seley, K. L.; Schneller, S. W.; Rattendi, D.; Bacchi, C. J. J. Med. Chem. 1997, 40, 622–624.
- For example, (a) Siddiqi, S. M.; Chen, X.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1994, 37, 551–554; (b) Tuncbilek, M.; Schneller, S. W. Bioorg. Med. Chem. 2003, 11,

- 3331-3334; (c) Roy, A.; Schneller, S. W.; Keith, K. A.; Hartline, C. B.; Kern, E. R. Bioorg. Med. Chem. 2005, 13, 4443-4449.
- 7. Yin, X. Q.; Schneller, S. W. Tetrahedron Lett. 2005, 11, 1927-1929.
- 8. Matsuumi, M.; Ito, M.; Kobayashi, Y. Synlett 2002, 1508– 1510.
- 9. (a) Rajappan, V. P.; Yin, X. Q.; Schneller, S. W. Tetrahedron 2002, 58, 9889–9895; (b) Yang, M.; Ye, W.; Schneller, S. W. J. Org. Chem. 2004, 69, 3993-3996.
- 10. Madhavan, G. V. B.; Martin, J. C. J. Org. Chem. 1986, 51,
- 1287–1293. 11. For 3: $[\alpha]_D^{23.4}$ –54.4 (*c* 0.149, DMSO). 12. Nakashima, H.; Sato, M.; Taniguchi, T.; Ogasawara, K. Synthesis 2000, 6, 817-823.
- 13. Rajappan, V. P.; Hegde, V. R.; Schneller, S. W. Syn. Commun. 2001, 31, 2849-2854.

- 14. Patil, S. D.; Schneller, S. W.; Hosoya, M.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. J. Med. Chem.
- 1992, 35, 3372–3377. 15. Data for 4: $[\alpha]_D^{23.4}$ +66.3 (c 0.135, DMSO); ¹H NMR (DMSO, 400 Hz) δ 8.24 (s, 1H), 8.18 (s, 1H), 7.31 (br s, 2H), 5.71 (d, J = 7.1 Hz, 1H), 5.42 (d, J = 8.3 Hz, 1H), 5.24 (d, J = 6.2 Hz, 1H), 4.90 (m, 2H), 4.71 (m, 1H), 4.14(dd, J = 12.8, 4.8 Hz, 1H), 3.99 (s, 1H); ¹³C NMR (DMSO, 62.5 Hz) δ 155.9, 152.4, 149.9, 140.1, 118.4, 92.1 (d, J = 187.0 Hz, 1C), 75.8 (d, J = 17.0 Hz, 1C), 73.7, 72.4, 57.6 (d, J = 15.4 Hz, 1C); ¹⁹F (DMSO, 250 Hz) -211.4 (m, 1F). Anal. Calcd for C₁₀H₁₂FN₅O₃: C, 44.61; H, 4.49; N, 26.01. Found: C, 44.49; H, 4.58; N, 25.72.
- 16. For **5**: $[\alpha]_D^{23.2}$ –16.5 (*c* 0.025, DMSO). 17. Yin, X. Q.; Rajappan, V. P.; Roy, A.; Schneller, S. W. Syn. Commun. 2003, 33, 1477–1481.